

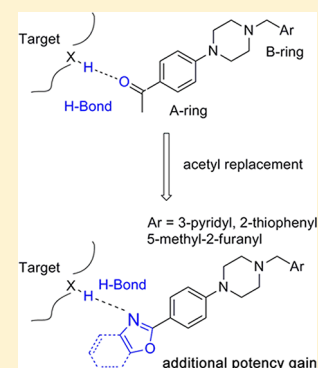
Discovery and Preliminary Structure–Activity Relationship of Arylpiperazines as Novel, Brain-Penetrant Antiprion Compounds

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Supporting Information

ABSTRACT: Creutzfeldt-Jakob disease and kuru in humans, BSE in cattle, and scrapie in sheep are fatal neurodegenerative disorders. Such illnesses are caused by the conversion and accumulation of a misfolded pathogenic isoform (termed PrP^{Sc}) of a normally benign, host cellular protein, denoted PrP^C. We employed high-throughput screening enzyme-linked immunosorbent assays to evaluate compounds for their ability to reduce the level of PrP^{Sc} in Rocky Mountain Laboratory prion-infected mouse neuroblastoma cells (ScN2a-cl3). Arylpiperazines were among the active compounds identified, but the initial hits suffered from low potency and poor drug-likeness. The best of those hits, such as **1**, **7**, **13**, and **19**, displayed moderate antiprion activity with EC₅₀ values in the micromolar range. Key analogues were designed and synthesized on the basis of the structure–activity relationship, with analogues **41**, **44**, **46**, and **47** found to have submicromolar potency. Analogues **41** and **44** were able to penetrate the blood–brain barrier and achieved excellent drug concentrations in the brains of mice after oral dosing. These compounds represent good starting points for further lead optimization in our pursuit of potential drug candidates for the treatment of prion diseases.

KEYWORDS: Neurodegenerative diseases, prion disease, Creutzfeldt-Jakob disease, piperazine, arylpiperazine, structure–activity relationship



Prion diseases are transmissible, rapidly progressive, fatal, and untreatable neurodegenerative disorders that afflict both humans and animals.^{1–3} An early key pathogenic event in diseases caused by the prion protein (PrP) is the conversion of the host cellular PrP, denoted PrP^C, into an aberrant disease-causing isoform, PrP^{Sc}, which accumulates in the central nervous system (CNS) of the infected host.^{4–6} The exact mechanism for the conversion of PrP^C to PrP^{Sc} remains unknown.

Currently, no treatments exist for prion diseases. Several classes of antiprion compounds that target levels of PrP^{Sc} or PrP^C, including approved drugs (for other uses) in humans as well as experimental small molecules, have been reported.^{7,8} Some FDA-approved drugs, including quinacrine, phenothiazines, and statins, showed antiprion activity in cell culture but demonstrated no in vivo efficacy when tested in prion-infected mice.^{9–11}

We have sought a strategy to treat prion disease by lowering the levels of PrP^{Sc} in prion-infected cell models. Reduced PrP^{Sc} levels are correlated with decreased prion infectivity in cultured cells and delayed development of clinical signs. Using HTS, we identified small molecules that lowered the levels of PrP^{Sc} in an RML prion-infected murine neuroblastoma cell line (ScN2a-cl3),¹² among which were the arylpiperazines (B. M. Silber et al., manuscript in preparation). Herein, we report SAR analysis and preliminary lead optimization to improve the potency and drug-likeness of the arylpiperazines and describe the discovery

of potent, metabolically stable, BBB-penetrant leads from this series. These compounds are excellent starting points for further lead optimization, with the goal of identifying candidates for further testing in scrapie- and CJD-infected animal models of prion disease.

From our HTS screening efforts of 53000 compounds, we identified 881 arylpiperazine analogues, 108 of which reduced PrP^{Sc} levels in ScN2a-cl3 cells by $\geq 30\%$. Of these 108 actives, 58 were subjected to a confirmatory assay and 41 of them were confirmed, i.e., reduced the level of PrP^{Sc} by $\geq 30\%$ without affecting cell viability ($\leq 30\%$ inhibition). A majority ($n = 29$) of these compounds, including the top 22 actives, are derivatives of *N*-arylpiperazine having a *para* methylketone or acetyl group on the phenyl ring (**1**, **7**, **13**, **14**, **18**, and **19**) (Table 1). Compounds bearing other substituents on the *N*-phenyl ring, regardless of whether they are electron-rich (**6**, **12**, **17**, and **22**) or electronic-deficient (**5**, **11**, **16**, and **21**) or their positions on the *N*-phenyl ring (compare **3** and **4**, **9** and **10**, **15** and **16**, and **20** and **21**), were found to be inactive under our assay conditions (Table 1). This finding argues for the importance of the 4-acetyl group of the *N*-aryl ring in determining the potency of those compounds. We initially postulated that the oxygen of the acetyl group is involved in a hydrogen bond (HB) with the

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Table 1. PrP^{Sc} Reduction for Selected Arylpiperazine Analogues

compd	R ₁	X	PrP ^{Sc} reduction (%)	
1	4-MeCO-	CH	94	
2	H	CH	5	
3	3-Cl	CH	7	
4	4-Cl	CH	6	
5	4-F	CH	7	
6	4-MeO	CH	7	
7	4-MeCO-	N	73	
8	H	N	4	
9	3-Cl	N	4	
10	4-Cl	N	0	
11	4-F	N	27	
12	4-MeO	N	10	

compd	R ₂	Y	R ₃	PrP ^{Sc} reduction (%)
13	4-MeCO-	S	H	91
14	4-MeCO-	S	4-Me	78
15	3-Cl	S	3-Br	34
16	4-Cl	S	2-Br	12
17	4-MeO	S	H	16
18	4-MeCO-	O	H	96
19	4-MeCO-	O	2-Me	78
20	3-Cl	O	H	6
21	4-Cl	O	H	13
22	4-MeO	O	2-Br	6

putative target protein (Figure 1a). Conversely, the substitution requirement on the N' atom of the piperazine is less stringent, and both arylmethylenes and heteroarylmethylenes, including benzyl (**1**), 3-pyridinylmethylene (**7**), 2-furanylmethylene (**19**),

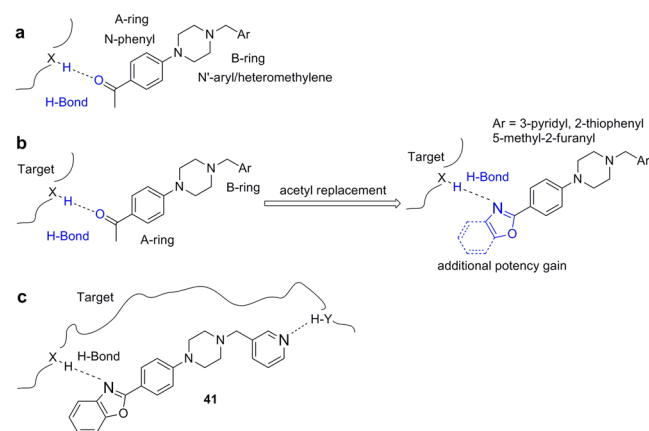


Figure 1. (a) Arylpiperazine lead, showing HB interaction between its 4-acetyl group and the putative target. (b) Design of piperazine analogues, via replacement of acetyl with oxazole or benzoxazole. (c) Compound **41**, showing its HB interactions with the putative target.

and 2-thiophenylmethylene (**13** and **14**), are well tolerated and show similar antiprion activities (Table 1).

In our A-ring SAR exploration, we initially tested the requirement for the 4-acetyl group. Compounds with various substituents, including 4-acetyl, chloro, fluoro, and methoxy as well as their regioisomers [**11**, **12**, and **23–26** (Table 1 of the Supporting Information)], were purchased and tested. All six compounds showed $\leq 30\%$ reduced PrP^{Sc} levels and EC₅₀ values of $>32 \mu\text{M}$; thus, they were considered inactive in our assays.

On the B-ring side, several examples of different (hetero)aryls representing phenyl (**1**), 3-pyridyl (**7**), thiophenyl (**13**), and furanyl (**19**) scaffolds were selected to test the influence of the B-ring on antiprion activity. Compounds **1**, **7**, **13**, and **19** were found to possess only moderate activities, with EC₅₀ values ranging from 0.71 to 4.62 μM (Supporting Information, Table 1). For our lead compounds, we aimed for >10 -fold potency compared to those identified in the ScN2a-cl3 cell assay. Thus, we embarked on an SAR effort to improve the antiprion potency of the arylpiperazine series. To ensure the decrease in the PrP^{Sc} level was not due to cytotoxicity, cell viability in ScN2a-cl3 cells was also determined to accompany each EC₅₀ enzyme-linked immunosorbent assay measurement using the fluorescent probe calcein-AM, as reported previously.¹³

We noted from our early SAR that the hydrogen bond acceptor (HBA) acetyl group is clearly important for antiprion potency of these analogues. We sought to replace it with alternative functionality that could preserve the HB interaction with the putative target. Initially, we surmised that a five-membered heterocycle, such as a 1,3-oxazole, might be a good first choice as an acetyl isostere. Both nitrogen and oxygen atoms of the oxazole are positioned optimally and are capable of functioning as an HBA. We also synthesized and examined the effect of fused benzoxazole systems for possible increased potency (Figure 1b). While the nitrogen or oxygen of the benzoxazole ring may function as an HBA, the fused phenyl ring may also provide additional binding affinity.

To ensure adequate coverage on the B-ring side, three aryl and heteroaryl rings, including 3-pyridine, 2-thiophene, and 5-methyl-2-furan, were selected for these initial SAR studies. A total of three oxazoles (**38–40**) and three benzoxazoles (**41–43**) were synthesized (Table 2 and Schemes 1 and 2). Almost without exception, we found that arylpiperazine analogues did not affect cell viability in ScN2a-cl3 cells (Table 2).

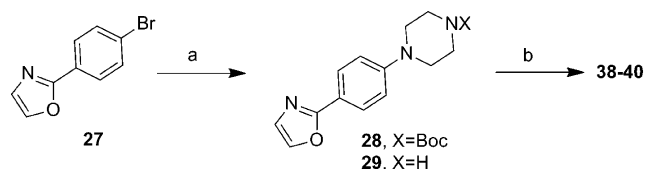
With respect to antiprion potency, oxazoles **38–40** were found to be only weakly active, with EC₅₀ values of $\geq 10 \mu\text{M}$, suggesting that a simple replacement of the HBA group by an oxazole ring was not sufficient to maintain potency. The additional phenyl ring, as seen in benzoxazoles **41–43** (Table 2), was beneficial to antiprion activity: all three analogues showed EC₅₀ values of $<10 \mu\text{M}$. The 3-pyridyl analogue **41** had an EC₅₀ of 0.25 μM , which was ≥ 13 -fold more potent than both furanyl **42** and thiophenyl **43** (Table 2). While we cannot specify the exact reason for such a potency increase for **41** versus those of other similar analogues with different B-rings, the pyridyl nitrogen atom in **41** may be situated to establish a second HB with the putative molecular target, contributing to its antiprion activity (Figure 1c).

To evaluate the BBB permeability of **41** and its potential as a CNS drug lead, we administered it to mice using a previously reported pharmacokinetic (PK) protocol.¹⁴ Rather than creating a full PK profile, we utilized an abbreviated protocol

Table 2. Antiprion Potencies (EC_{50}) and Cell Viabilities (LC_{50}) for Selected Arylpiperazine Analogues

compd	R ¹	R ²	$EC_{50} \pm SEM$ (μM) ^a	LC_{50} (μM) ^a
38			>10	>10
39			>10	>10
40			>10	>10
41			0.3 ± 0.0	>10
42			3.3 ± 0.5	>10
43			4.2 ± 2.2	>10
44			0.4 ± 0.0	>10
45			>10	>10
46			0.4 ± 0.0	>10
47			1.1 ± 0.6	>10

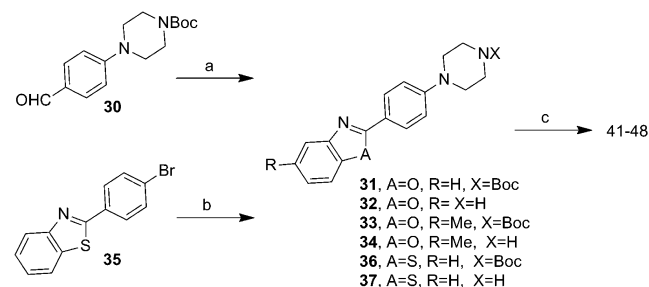
^aThree tests for EC_{50} and LC_{50} measurements.

Scheme 1. Synthesis of Oxazole Analogues 38–40^a

^aReaction conditions: (a) (i) *tert*-butyl piperazinecarboxylate/*t*-BuOK, Pd₂(dba)₃ (5% mmol), BINAP (15% mmol), toluene, reflux, 12 h, 70%; (ii) TFA, DCM, room temperature, 3 h, 93%; (b) RCHO/HCOOH (2 equiv), anhydrous DMF, 100 °C, 3 h, 21–37%.

to determine key parameters, including C_{max} and AUC, which are important for initial compound selection and prioritization. For initial PK studies, all compounds were administered in a single dose (10 mg/kg) by oral gavage in a formulation containing 20% propylene glycol, 5% ethanol, 5% labrosol, and 70% PEG400. Brain and plasma concentrations were measured at 0.5, 2, 4, and 6 h after administration. The C_{max} and the AUC from 0 to 6 h (AUC_{0-6h}) for both brain and plasma were calculated. Compound **41** exhibited an excellent brain concentration [$C_{max} = 3.63 \mu M$ (Table 3)]. Its very good brain exposure ($AUC_{0-6h} = 14.2 \mu M h$) was approximately 4-fold higher than its plasma exposure ($AUC_{0-6h} = 4.18 \mu M h$), a desired characteristic in a CNS drug. Compound **41** was better absorbed considering its poor microsomal stability [$t_{1/2} = 9.2$ min (Table 3)].

Encouraged by the *in vitro* antiprion potency and *in vivo* PK profile, we decided to expand the SAR around lead **41**, hoping to identify more potent analogues. On the A-ring side, we

Scheme 2. Synthesis of Compounds 41–48^a

^aReaction conditions: (a) (i) 2-aminophenol or 2-amino-4-methylphenol, O₂, TEMPO, toluene, reflux overnight, 23–81%; (ii) TFA/DCM, room temperature, 3 h, 66–77%; (b) (i) *tert*-butyl piperazinecarboxylate/*t*-BuOK, Pd₂(dba)₃ (5% mmol), BINAP (15% mmol), toluene, reflux, 12 h, 60%; (ii) TFA/DCM, 25 °C, 3 h, 85%; (c) RCHO/HCOOH (2 equiv), anhydrous DMF, 100 °C, 3 h, 6–44%.

synthesized a methylated benzoxazole [**48** (Scheme 2)] to probe the size constraints of the A-ring binding pocket. Compound **48** was less potent than **41**, with an EC_{50} of 1.65 μM , indicating that substitution of benzoxazole with a methyl group (**48**) was not well-tolerated (data not shown).

We also examined the effect of hydrophobicity on binding affinity by replacing the benzoxazole ring (**41**) with a more hydrophobic benzothiazole ring [**44** (Scheme 2)]. The sulfur atom of **44** is not capable of acting as an HBA, a role assumed by the oxygen atom in the benzoxazole **41**; the outcome of this compound may allow us to determine which atom (nitrogen or oxygen) in **41** is involved in the HB with the putative target. Replacing the oxygen atom in **41** with a sulfur atom (**44**) had little impact on antiprion activity: **44** was slightly less potent than **41**, with an EC_{50} of 0.38 μM (Table 2). Presumably, **44** still maintains a HB with the putative target via the nitrogen atom of the benzothiazole ring.

On the B-ring side, we believe the nitrogen atom of **41** is involved as an HBA with the putative target. Addition of a substituent next to the nitrogen may perturb this interaction and thus influence the potency of these analogues. Additionally, because the nitrogen in the pyridine ring is often a target for cytochrome P450 (CYP450) enzymes, the introduction of an adjacent group, in addition to modulating the HB potential and its potency, may also improve the metabolic stability of these pyridine analogues. Therefore, we designed and synthesized several analogues with substituents (**45–47**) next to the nitrogen atom of the pyridine ring (Scheme 2).

4-Methyl analogue **45** was found to be inactive, with an EC_{50} value of >10 μM (Table 2); the B-ring binding pocket should be wide enough to accommodate a small methyl group based on an early SAR derived from phenyl, thiophene, and furan analogues. As a result, the loss of the antiprion potency of **45** may be due to interference of HB formation between the adjacent nitrogen and its target. In contrast, both 4-F and 4-MeO analogues (**46** and **47**, respectively) were active, with EC_{50} values of 0.38 and 1.11 μM , respectively (Table 2). 4-Fluoropyridine analogue **46** was 1.5-fold less potent than its parent pyridine analogue **41**. This observation might be explained by a reduction in the strength of the HB between the nitrogen and the putative target; the neighboring fluoro atom reduces the electron density on the nitrogen atom and thus weakens the strength of the HB. 4-MeO analogue **47** was

Table 3. In vivo PK Parameters and in vitro Microsomal Stability for Arylpiperazines (single oral 10 mg/kg dose)

compd	brain exposure		plasma exposure		microsomal stability, $t_{1/2}$ (min)	
	C_{\max} (μM)	AUC ($\mu\text{M h}$)	C_{\max} (μM)	AUC ($\mu\text{M h}$)	Mo	Hu
41	3.6 \pm 0.9	14.2 \pm 0.6	0.9 \pm 0.1	4.2 \pm 0.6	9.2	50.2
44	3.0 \pm 0.5	14.4 \pm 0.2	0.6 \pm 0.0	2.7 \pm 0.4	17.3	34.3
46	0.4 \pm 0.1	1.3 \pm 0.2	0.4 \pm 0.2	1.2 \pm 0.1	7.3	43.6
47	1.6 \pm 0.8	3.0 \pm 1.0	0.7 \pm 0.4	1.1 \pm 0.5	25.2	>60

4.5-fold less potent than **41**. The larger methoxy group may hinder and weaken the HB forming ability or strength of the nitrogen with the target.

Next, we tested the metabolic stability of compounds **44**, **46**, and **47** in mouse and human microsomes (Table 3). Compared to benzoxazole **41**, benzothiazole **44** had approximately 2-fold improvement ($t_{1/2}$ = 17.3 and 9.2 min for **44** and **41**, respectively) in mouse microsomes but was less stable in human microsomes ($t_{1/2}$ = 34.3 and 50.2 min for **44** and **41**, respectively). 4-Fluoro analogue **46** was less stable in both mouse and human microsomes than **41**. The fluorine atom could affect the oxidation of the pyridine ring, but it also increases the overall hydrophilicity (ChemDraw calculated logP values of 3.86 and 4.09 for **41** and **46**, respectively) of the molecule and may open the possibility of oxidation in other parts of the molecule. Methoxy analogue **47** had an approximately 3-fold increase in stability in both human and mouse microsomes.

We then profiled compounds **44**, **46**, and **47** via in vivo PK studies as described above, focusing on their BBB permeability and brain exposure (Table 3). Both B-ring analogues **46** and **47** had poor exposure in brain and plasma compared to their parent pyridine analogue **41**. Benzothiazole analogue **44**, however, displayed a favorable PK profile compared to that of **41**, with a 5-fold brain:plasma exposure ratio.

In summary, the initial arylpiperazine HTS hits with low micromolar activity (2.94 μM for **7**) were optimized to yield several leads with a >10-fold antiprion potency improvement. The acetyl group in the original HTS hits was replaced with the more druglike benzoxazole ring. These efforts culminated with the identification of **41** and **44**, both demonstrating robust antiprion potency, favorable PK properties, and good CNS penetration. Further evaluation of these compounds in RML- and CJD-infected animal models is underway.

■ ASSOCIATED CONTENT

Supporting Information

Table 1, experimental procedures, and synthetic procedures for all intermediates. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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All authors have given approval to the final version of the manuscript.

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Notes

The authors declare no competing financial interest.

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■ ABBREVIATIONS

BBB, blood-brain barrier; BINAP, 2,2'-bis-(diphenylphosphino)-1,1'-binaphthyl; CJD, Creutzfeldt-Jakob disease; DCM, dichloromethane; EC₅₀, half-maximal effective concentration; HBA, hydrogen bond acceptor; HBD, hydrogen bond donor; HTS, high-throughput screening; Pd₂(dba)₃, tris(dibenzylideneacetone)dipalladium(0); PrP^C, normal cellular isoform of the prion protein; PrP^{Sc}, pathogenic isoform of the prion protein; RML, Rocky Mountain Laboratory; SAR, structure-activity relationship; TEMPO, (2,2,6,6-tetramethylpiperidin-1-yl)oxyl or (2,2,6,6-tetramethylpiperidin-1-yl)-oxidanyl; TFA, trifluoroacetic acid.

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